Original Article

Role of Fatty Acids Intake in Generalized Vitiligo

Abstract

Background: Previous studies indicated the effect of fat on autoimmune diseases. The present study was aimed to investigate the association between fat intake and vitiligo. Methods: This case-control study was conducted in the Skin and Leishmania Research Center, Isfahan University of Medical Sciences, Isfahan, Iran. Intakes of fatty acids were examined for their relation to risk of vitiligo among 100 cases and 110 controls. We included patients who suffered from generalized or localized vitiligo for <5 years that was approved by a dermatologist via the Vitiligo European Task Force criteria and the vitiligo area scoring index. Fat intake was assessed through individual interviews by a standardized food frequency questionnaire. Results: Vitiligo group consumed more saturated fatty acid (SFA) and less eicosapentaenoic acid and docosahexaenoic acid than control group, while other fatty acids were not significantly different among two groups (P > 0.05). Crude analysis showed that total fat (odds ratio [OR] = 3.33, 95% confidence interval [CI]: 1.46–7.58) and SFA (OR = 2.22, 95% CI: 1.04-4.90) intakes were associated with an increased risk of vitiligo (for highest quartile vs. lowest quartile). Results demonstrated a decrease in the risk of vitiligo for those within the highest quartile of monounsaturated fatty acids intake (OR = 0.41, 95% CI: 0.18-0.92). However, this relationship disappeared after adjustment for confounders as energy, age, sex, and body mass index, except for total fat (OR = 2.84, 95% CI: 1.63-5.44). Crude and adjusted analyses for polyunsaturated fatty acids and cholesterol intake were not statistically significant. Conclusions: Total-fat content of the diet had more impressive role than the specific subclasses of fats on the incidence risk of vitiligo. High-fat diet escalated the vitiligo risk. Regarding the role of fats on skin autoimmune diseases especially vitiligo, future studies are crucial.

Keywords: Autoimmune diseases, fatty acids, vitiligo

Introduction

Vitiligo is one of the skin diseases that cause white spots due to loss of skin pigment cells, which is considered as an autoimmune disease.[1] Melanocytes, mucous membrane, and retina are damaged and lead to white spots in different areas of the skin.[2] The face, lips, hands, arms, feet, and the genitals are commonly affected skin area. Moreover, the color of the hairs in affected areas is usually white.[3] The prevalence of vitiligo is various between 0.38% and 2.9% in worldwide that affect all races and two genders. Other autoimmune disorders enhance prevalence of this disease. [2,4,5] The exact etiopathogenic mechanism of vitiligo is not understood.[6] Loss of melanocytes may be associated with autoimmune cytotoxic T-cells. oxidant-antioxidant imbalance. genetic factors, neural mechanisms, or multifactorial mechanisms.[7-10] Cellular

important effect on the modulation of the immune system. It has been demonstrated in the numerous experimental studies. [13] Therefore, a strong relationship between dietary fat and modulation of immune response could be established. Several studies revealed the importance of fatty

autoimmunity plays a major role in vitiligo

Functions of immune system may be

influenced by the nutritional status; lipids

as crucial components in diets have

pathogenesis.[11,12]

response could be established. Several studies revealed the importance of fatty acids in the diet and their application on the reduction of typical symptomatologies in autoimmune diseases. [14,15] Dietary fat has different effects on immune system based on the type of fat. N-3 fatty acids that commonly exist in fish oil have shown a significant reduction of inflammation in patients suffering from rheumatoid arthritis.

Other findings have indicated that different fats such as olive oil, oleic acid, eicosa

How to cite this article: Derakhshande-Rishehri S, Heidari-Beni M, Jaffary F, Askari G, Nilfroshzade M, Ansari N, *et al.* Role of fatty acids intake in generalized vitiligo. Int J Prev Med 2019;10:52.

psoriasis, systemic lupus erythematosus, multiple sclerosis (MS), etc.^[16-19]
Other findings have indicated that different

Seyedeh-Masomeh Derakhshandeh-Rishehri^{1,6}, Motahar Heidari-Beni², Fariba Jaffary³, Gholamreza Askari⁴, Mohammadali Nilfroshzade^{5,6}, Neda Adibi³

¹Food Security Research Center, Department of Community Nutrition, School of Nutrition and Food Sciences, Isfahan University of Medical Sciences, Isfahan, Iran, ²Child Growth and Development Research Center, Research Institute for Primordial Prevention of Non-communicable Disease, Isfahan University of Medical Sciences, Isfahan, Iran, 3Skin Diseases and Leishmaniasis Research Center, Isfahan University of Medical Sciences, Isfahan, Iran, ⁴Department of Community Nutrition, Food Security Research Center, School of Nutrition and Food Sciences, Isfahan University of Medical Sciences, Isfahan, Iran, ⁵Skin and Leishmania Research Center, Isfahan University of Medical Sciences, Isfahan, Iran, ⁶Skin and Stem Cell Research Center, Tehran University of Medical Sciences, Tehran, Iran

Address for correspondence:
Dr. Fariba Jaffary,
Skin Diseases and Leishmaniasis
Research Center, Isfahan
University of Medical Sciences,
Isfahan, Iran.
E-mail: jaffary@pharm.mui.ac.ir

A ---- Alais autiala autias

Access this article online

Website:

www.ijpvmjournal.net/www.ijpm.ir

10.4103/ijpvm.lJPVM_47_17

Quick Response Code:



This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

pentaenoic (EPA), or docosahexaenoic (DHA) acids modulated the function of immune system. [20-23] However, according to some findings, polyunsaturated fatty acids (PUFAs) had more immunosuppressive effect than saturated fatty acids (SFA). Others showed that dietary PUFAs may interfere in the reduction of lymphocyte proliferation, cytokine synthesis, natural killer cell (NK) activity, antibody production, membrane surface molecules synthesis, etc. [24-28]

Findings about the association between dietary fat and autoimmune disease are controversial. Studies on dietary intake and vitiligo are less and investigation in this regard is necessary. The aim of the present study was investigation the association between fat intake and vitiligo.

Methods

Participants

A total of 147 vitiligo patients referred to Skin and Leishmania Research Center, Isfahan University of Medical Sciences, Isfahan, Iran. In this case-control study, we recruited 100 vitiligo patients and 110 healthy subjects among eligible volunteers. We included individuals who those suffered from generalized or localized vitiligo for <5 years after their condition was approved by a center dermatologist. Other inclusion criteria were no positive family history of vitiligo, no use of systemic or topical treatment, anti-inflammatory or immunosuppressive drugs, corticosteroids, augmenter or depressor of lipid profile for at least 1 month before the study, no concomitant dermatological or systemic diseases, lack of underlying illnesses which demand special diet such as diabetes, hypertension, or renal disease, lack of heavy exercises, special diet, smoking, pregnancy, or lactation. The exclusion criteria were pregnancy or lactation during the study, diagnosis of disease such as bacterial or viral infections, or acute illnesses and start drug therapy during the study, mental disability to fill out the questionnaire, and lack of interest to continue cooperation.

General characteristic survey

General characteristics including sex, age, smoking, medical history (medication use and history of diseases such as diabetes, cancer, and cardiovascular disease), physical activity, anthropometric data (weight, height, and body mass index [BMI]), presence of food limitation, and list of restricted foods were obtained by questionnaire.

The list of restricted foods in the present study included eggs, milk and dairy products (milk, cheese, plain yogurt), grains (wheat), fish (white flesh fish, red flesh fish), meats (beef, chicken), oily or spicy foods (soda, food additives, tinned foods or drinks, sour or pickled food items, tamarind,), citrus fruits and juices, grapes, pears, tomatoes, cherries, mangoes, chocolate and cocoa products, coffee, and others which could possibly aggravate vitiligo

patches of depigmentation areas. Furthermore, the list of drugs was as follows: steroids, Ultraviolet B Light (either broadband or narrowband), psoralen and ultraviolet A light, tacrolimus (immunosuppressive), pimecrolimus (immunomodulator), skin camouflage solutions, depigmenting drugs such as (monobenzone, mequinol, or hydroquinone). All participants signed the consent form consciously; furthermore, patients were allowed to withdraw the study at any time that they were not willing to continue working with the team.

Vitiligo severity indices and treatment evaluation criteria

Over the past decade, two methods of evaluation, while differ in their approach and outcomes, have been recognized as validated standards for the comparative evaluation of vitiligo and vitiligo treatments under clinical conditions: (1) the vitiligo area scoring index and (2) the Vitiligo European Task Force (VETF) system. Similarly, in both assessments, the body is separated into five different sites, specifically the head/neck, trunk, arms, legs, and hands/feet. In the present study, the center dermatologist used VETF as diagnostic method. The VETF evaluation system seeks to add more specific parameters to the quantitative measurement of depigmentation. Indeed, the VETF assesses the three dimensions of the disease (extent, staging, and spreading/progression). In VETF method, in each site, the largest lesion within each specific body site is clinically evaluated by visual and photographic assessment for the extent or percentage of vitiligo involvement (depigmented skin), staging, and spreading of vitiligo. In VETF method, staging is assessed from 0 (normal pigmentation) to 4 (complete hair whitening) grades. Spreading is assessed using the following scores: 0 (stable disease), -1 (regressive disease), and +1 (progressive disease).

Dietary assessments

Each participant's usual food intake over the previous year was obtained through individual interviews by standardized food frequency questionnaire (FFQ) that contained 136 food items. Validity and reliability of questionnaire were approved. [29,30] Administered interviewer collected dietary information via face-to-face interview. All of the foods that were consumed at least twice a month were registered. The portions of each consumed food (g/d) were quantified by household measures, standard measures, and 35 sets of pictures with simple foods, food mixtures, and drinks. Ouestionnaire had an open-ended section where respondents recorded consumption of other foods not included on the food list and we ensured that the total diet of the individual was captured. In addition, FFQs include supplementary questions about cooking methods and specific types of fat and milk. A food composition table was used for mixed foods ingredients. Food items were converted to gram. Nutritionist 4 software (First Data Bank, San Bruno, CA) was used for nutrient analysis.

Statistical analysis

Results were expressed as percentage (qualitative variables) and mean ± standard deviation (quantitative variables). Dietary total fat, PUFA, monounsaturated fatty acids (MUFA), SFA, linoleic acid, linolenic acid, oleic acid, EPA, DHA, and cholesterol variables were adjusted for total energy intake using the residual method. Independent t-test was used to compare the amount of total fat, PUFA, MUFA, SFA, linoleic acid, linolenic acid, oleic acid, EPA, DHA, and cholesterol intake between two groups. Multiple logistic regression models were used to analyze the association between dietary intake of fatty acids and occurrence of vitiligo with adjusting of potential confounders (age, sex, BMI, physical activity, and energy). Odds ratios (ORs) and corresponding 95% confidence intervals (CIs) for the risk of vitiligo at different quartiles of dietary total fat, PUFA, MUFA, SFA, and cholesterol intake were computed in unadjusted and adjusted analyses. Quartile cut-points were based on the distribution of variables among controls. All analyses were performed with SPSS version 20.0 (SPSS, Chicago, IL, USA). Results with a value of P < 0.05 were considered statistically significant.

Results

Data of 100 vitiligo patients (67 women, 38 men) and 110 healthy volunteers (33 women, 62 men) were analyzed. We found no significant differences between the cases and the controls in terms of age, sex, and BMI, but there was a significant difference in weight (P = 0.009) [Table 1].

Results from the independent *t*-test showed that consumption of SFA, EPA, and DHA significantly differed between the cases and the controls. Vitiligo group intakes more SFA and less EPA and DHA than control group. The intakes of energy, total fatty acids, MUFAs, PUFAs, cholesterol, oleic acids, linoleic acids, and linolenic acids were not significantly differed among two groups [Table 2].

Table 3 presents the results of the multiple logistic regression models regarding the relationship between quartiles of fatty acids intake and risk of vitiligo as ORs, 95% CIs, and *P* values for trend.

According to Table 3, crude analysis showed increase in the risk for those with high intakes of total fat (OR = 3.33, 95% CI: 1.46–7.58) and SFA (OR = 2.22, 95% CI: 1.04–4.90) and decrease in the risk for those with high intakes of MUFA (OR = 0.41, 95% CI: 0.18–0.92). However, this relationship disappeared after adjustment for confounders such as energy, age, sex, and BMI except for total fat (OR = 2.84, 95% CI: 1.63–5.44). Crude and adjusted analyses for PUFA and cholesterol intake were not statistically significant [Table 3].

Discussion

In the present case-control study, analysis confirmed that higher total fat intake associated with higher incidence risk

Table 1: Characteristics of participants				
	Vitiligo	Control	P	
Age	20.71±6.19	23.22±5.39	0.07	
Gender (%)				
Male	38	62	0.052	
Female	67	33		
Weight	57.31±8.9	65.87±2.66	0.009	
BMI (kg/m²)	21.48±2.68	23.31±5.86	0.051	

BMI=Body mass index

Table 2: Dietary fat intake in two groups of study							
	Control (n=110)	Vitiligo (n=100)	P				
Energy	2453.93±984.10	2147.35±874.38	0.21				
Total fat	71.37 ± 20.46	76.87 ± 19.72	0.053				
SFA	22.41±7.52	25.35 ± 8.14	0.008				
MUFA	23.81 ± 14.78	22.39 ± 12.27	0.46				
PUFA	18.13±8.60	17.90 ± 6.80	0.08				
Linoleic acid	16.27±11.57	14.25 ± 9.24	0.17				
Linolenic acid	0.42 ± 0.32	0.35 ± 0.25	0.11				
Oleic acid	18.4±13.25	16.43 ± 9.83	0.23				
EPA	0.03 ± 0.04	0.01 ± 0.01	0.001				
DHA	0.07 ± 0.12	0.04 ± 0.03	0.004				
Cholesterol	229.36±88.77	252.31±120.64	0.82				

SFA=Saturated fatty acid, MUFA=Monounsaturated fatty acid, PUFA=Polyunsaturated fatty acid, EPA=Eicosapentaenoic acid, DHA=Docosahexaenoic acid

of vitiligo. Although an unfavorable association was shown between the higher intake of SFA and lower intakes of EPA or DHA and vitiligo, these associations disappeared after adjustment with confounders.

Limited studies assessed the relationship between dietary fat intakes and the incidence or severity of vitiligo.

In clinical studies, significant benefits for systemic lupus erythematosus patients were achieved after consuming a low-fat diet plus n-3 fatty acid-rich fish oil supplement. [18] Studies showed that high saturated fat diets have deleterious effects on both macrophage phagocytosis and NK cell activity in autoimmune disease. [31] Autoimmune model study reported that a high-fat diet consisting of equal amounts of lard and soybean oil (rich in linoleic acid) developed disease and animals had a shortened lifespan. [32,33]

The composition of dietary fatty acids influenced the tissue phospholipids which in turn determine the amounts and types of precursor acids eicosanoids. There are two ways in which dietary fatty acids can modulate the biosynthesis of eicosanoids from arachidonic acid, the major 20-carbon PUFAs of the human monocytes and lymphocytes: (1) Essential fatty acids deficiency and high levels of trans-isomers of linoleic acid in the diet decrease tissue arachidonic acid and the biosynthesis of eicosanoids derived from arachidonic acid. (2) Dietary PUFA can modulate the biosynthesis of eicosanoids via the cyclooxygenase step.^[34,35] It has been speculated that

	Table 3: Multiple logistic regression and 95% confidence interval across quartile of fat intake						
	Q1	Q2	Q3	Q4	P trend		
Total fat							
Crude	1	2.33 (1.03-5.26)	1.91 (0.84-4.34)	3.33 (1.46-7.58)	0.01		
Adjusted	1	1.99 (1.21-2.45)	1.03 (0.54-3.55)	2.84 (1.63-5.44)			
SFA							
Crude	1	0.75 (0.28-1.47)	0.87 (0.54-1.61)	2.22 (1.04-4.90)	0.06		
Adjusted	1	0.27 (0.04-1.67)	0.37 (0.07-1.91)	0.59 (0.09-3.57)			
MUFA							
Crude	1	0.18 (0.07-0.43)	0.78 (0.35-1.73)	0.41 (0.18-0.92)	0.001		
Adjusted	1	0.14 (0.01-1.09)	0.51 (0.12-2.22)	0.23 (0.04-1.10)			
PUFA							
Crude	1	1.08 (0.49-2.35)	0.67 (0.30-1.47)	0.85 (0.39-1.86)	0.45		
Adjusted	1	0.99 (0.54-2.04)	0.53 (0.23-1.74)	0.79 (0.48-1.78)			
Cholesterol							
Crude	1	0.85 (0.38-1.86)	0.95 (0.43-1.23)	1.60 (0.73-3.50)	0.25		
Adjusted	1	0.75 (0.15-3.67)	0.82 (0.19-3.50)	0.88 (0.14-5.32)			

SFA=Saturated fatty acid, MUFA=Monounsaturated fatty acid, PUFA=Polyunsaturated fatty acid

changes in membrane fatty acid composition via dietary lipids can alter membrane fluidity, which in turn can change activities of antigen receptors, membrane-bound enzymes, and membrane permeability to ions, particularly Ca2⁺⁺.^[36,37]

Regarding the effects of SFAs on autoimmune system, Park *et al.*^[38] investigated the impact of high-fat diet (HFD), partially substituted with pine nut oil and lard for 12 weeks. They claimed that the production of IL-1 β by splenocytes was augmented in HFD mice; thus, IL-1 β triggers the immune responses. Furthermore, Jahromi *et al.*^[39] conducted a study to find the relation of dietary pattern and the risk of MS with factor analysis. They observed that traditional pattern high in low-fat dairy products and red meat was inversely associated with the risk of MS. Ghadirian *et al.*^[40] found that pork and hotdog intakes escalate the risk of MS. They also suggested a positive relation between energy and animal fat intake and the risk of MS.

Previous studies investigated the effect of omega-3 (ω3) PUFA on autoimmune diseases. Ghorbanibirgani et al.[41] indicated that Nigella sativa oil and fish oil reduced the size of vitiligo's lesions. In a study that was performed on 39 chronic psoriasis patients in Birjand, Iran, the fish oil had the same impact in reducing the size of skin lesions in comparison to a combination of salicylic acid and betamethasone.[14] In another clinical study, the positive effect of fish oil on skin autoimmune diseases such as vitiligo was confirmed in India.[42] Löfvenborg et al.[15] showed that fatty fish consumption might reduce the risk of latent autoimmune diabetes in adults, possibly through effects of ω3 fatty acids. Regarding the effects of ω3 fatty acids on the formation of eicosanoids from arachidonic acid, Lands et al.[43] demonstrated that n-3 PUFA competitively inhibits the oxygenation of arachidonic acid by cyclooxygenase. Hwang et al.[44] claimed that among different PUFAs, EPA (20:5 [n-3]) and DHA (22:6 [n-3]) are more effective than 18:3 (n-3) in suppressing tissue levels of arachidonic

acid and the formation of eicosanoids from arachidonic acid. On the other hand, Ochi *et al.*^[45] indicated that EPA was a poorer substrate for cyclooxygenase than arachidonic acid although it can be converted to thromboxane 3 and triene prostaglandins to a limited extent in tissues.

Some experimental studies assessed that the effect of ω6 PUFAs on autoimmune disorders showed sunflower oil, rich in linoleic acid, decreased relapse rate and severity of MS. [46,47] However, another study did not find this effect. [48]

Findings showed that high n-3 fatty acid diets increased the survival and reduced disease severity in spontaneous autoantibody-mediated disease, while linoleic acid-rich diets appear to increase disease severity. The underlying involved mechanisms were (1) regulation of gene expression, (2) signal transduction pathways, (3) production of eicosanoids and cytokines, (4) and the action of antioxidant enzymes.^[13]

Reasons for different results are the impact of dietary fatty acids on animal autoimmune disease models appears to depend on the animal model and the type and amount of fatty acids fed, effect of other environmental factor and genetic factors.

Vitiligo is considered as rare diseases or diseases with a long latency period between exposure and disease manifestation, and this is the first study that assesses the relation of dietary fats with vitiligo.

Conclusions

We conclude that the protective or detrimental effects of the dietary fatty acids on the risk of vitiligo are more dependent on the total fat content of the diet than the specific subclasses of fats or fatty acids. EPA and DHA may have beneficial effect on the treatment of vitiligo patients.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

Received: 21 Jan 17 Accepted: 19 Jan 18

Published: 06 May 19

References

- Rodríguez-Martín M, Sáez M, Merino de Paz N, Ferrer PC, Eliche MP, Rodríguez-Martín B, et al. When are laboratory tests indicated in patients with vitiligo? Dermatoendocrinol 2012;4:53-7.
- Burns T, Breathnach S, Cox N, Griffiths C. Rook's Textbook of Dermatology. Oxford: Blackwell Science; 2010. p. 164-6.
- James WD, Elston DM, Berger TG, Andrews GC. Andrews' Diseases of the Skin: Clinical Dermatology. Philadelphia: Saunders; 2011. p. 115-6.
- Nunes DH, Esser LM. Vitiligo epidemiological profile and the association with thyroid disease. An Bras Dermatol 2011;86:241-8.
- Bolognia JL, Jorizzo JL, Rapini R. Dermatology. Philadelphia: Mosby; 2008. p. 214.
- Nogueira LS, Zancanaro PC, Azambuja RD. Vitiligo and emotions. An Bras Dermatol 2009;84:41-5.
- Passeron T, Ortonne JP. Physiopathology and genetics of vitiligo. J Autoimmun 2005;25 Suppl:63-8.
- Dell'anna ML, Picardo M. A review and a new hypothesis for non-immunological pathogenetic mechanisms Pigment Cell Res 2006;19:406-11.
- Gauthier Y, Cario Andre M, Taïeb A. A critical appraisal of vitiligo etiologic theories. Is melanocyte loss a melanocytorrhagy? Pigment Cell Res 2003;16:322-32.
- Westerhof W, Manini P, Napolitano A, d'Ischia M. The haptenation theory of vitiligo and melanoma rejection: A close-up. Exp Dermatol 2011;20:92-6.
- Mandelcorn-Monson RL, Shear NH, Yau E, Sambhara S, Barber BH, Spaner D, et al. Cytotoxic T lymphocyte reactivity to gp100, melanA/MART-1, and tyrosinase, in HLA-A2-positive vitiligo patients. J Invest Dermatol 2003;121:550-6.
- Pichler R, Sfetsos K, Badics B, Gutenbrunner S, Berg J, Auböck J, et al. Lymphocyte imbalance in vitiligo patients indicated by elevated CD4+/CD8+T-cell ratio. Wien Med Wochenschr 2009;159:337-41.
- 13. Harbige LS. Dietary n-6 and n-3 fatty acids in immunity and autoimmune disease. Proc Nutr Soc 1998;57:555-62.
- Shahian Madar R, Ghaderi F. Comparison of acid salisylic 2% and betamethasone with topical fish oil in recovery of psoriasis signs. J Qazvin Univ Med Sci 2004;8:3-7.
- Löfvenborg JE, Andersson T, Carlsson PO, Dorkhan M, Groop L, Martinell M, et al. Fatty fish consumption and risk of latent autoimmune diabetes in adults. Nutr Diabetes 2014;4:e139.
- Kremer JM, Lawrence DA, Jubiz W, DiGiacomo R, Rynes R, Bartholomew LE, et al. Dietary fish oil and olive oil supplementation in patients with rheumatoid arthritis. Clinical and immunologic effects. Arthritis Rheum 1990;33:810-20.
- Bittiner SB, Tucker WF, Cartwright I, Bleehen SS. A double-blind, randomised, placebo-controlled trial of fish oil in psoriasis. Lancet 1988;1:378-80.
- Walton AJ, Snaith ML, Locniskar M, Cumberland AG, Morrow WJ, Isenberg DA, et al. Dietary fish oil and the severity

- of symptoms in patients with systemic lupus erythematosus. Ann Rheum Dis 1991;50:463-6.
- Bates D, Cartlidge NE, French JM, Jackson MJ, Nightingale S, Shaw DA, *et al.* A double-blind controlled trial of long chain n-3 polyunsaturated fatty acids in the treatment of multiple sclerosis. J Neurol Neurosurg Psychiatry 1989;52:18-22.
- Jeffery NM, Yaqoob P, Newsholme EA, Calder PC. The effects of olive oil upon rat serum lipid levels and lymphocyte functions appear to be due to oleic acid. Ann Nutr Metab 1996;40:71-80.
- De Pablo MA, Ortega E, Gallego AM, Alvarez C, Pancorbo PL, Alvarez de Cienfuegos G. Influence of diets containing olive oil, sunflower oil or hydrogenated coconut oil on the immune response of mice. J Clin Biochem Nutr 1998;25:11-23.
- de Pablo MA, Ortega E, Gallego AM, Alvarez C, Pancorbo PL, Alvarez de Cienfuegos G, et al. The effect of dietary fatty acid manipulation on phagocytic activity and cytokine production by peritoneal cells from Balb/c mice. J Nutr Sci Vitaminol (Tokyo) 1998;44:57-67.
- Calder PC, Yaqoob P, Harvey DJ, Watts A, Newsholme EA. Incorporation of fatty acids by concanavalin A-stimulated lymphocytes and the effect on fatty acid composition and membrane fluidity. Biochem J 1994;300(Pt 2):509-18.
- 24. Endres S, Ghorbani R, Kelley VE, Georgilis K, Lonnemann G, van der Meer JW, et al. The effect of dietary supplementation with n-3 polyunsaturated fatty acids on the synthesis of interleukin-1 and tumor necrosis factor by mononuclear cells. N Engl J Med 1989;320:265-71.
- 25. Yaqoob P, Newsholme EA, Calder PC. Inhibition of natural killer cell activity by dietary lipids. Immunol Lett 1994;41:241-7.
- Kelley DS, Taylor PC, Nelson GJ, Schmidt PC, Ferretti A, Erickson KL, et al. Docosahexaenoic acid ingestion inhibits natural killer cell activity and production of inflammatory mediators in young healthy men. Lipids 1999;34:317-24.
- 27. Lim BO, Yamada K, Hung P, Watanabe T, Taniguchi S, Sugano M, et al. Effects of n-3 polyunsaturated fatty acids and lectins on immunoglobulin production by spleen lymphocytes of Sprague-Dawley rats. Biosci Biotechnol Biochem 1996;60:1025-7.
- Sherrington EJ, Sanderson P, Calder PC. The effect of dietary lipid manipulation on macrophage cell surface molecule expression. Biochem Soc Trans 1995;23:272S.
- Mirmiran P, Esfahani FH, Mehrabi Y, Hedayati M, Azizi F. Reliability and relative validity of an FFQ for nutrients in the Tehran lipid and glucose study. Public Health Nutr 2010;13:654-62.
- 30. Malekshah AF, Kimiagar M, Saadatian-Elahi M, Pourshams A, Nouraie M, Goglani G, et al. Validity and reliability of a new food frequency questionnaire compared to 24 h recalls and biochemical measurements: Pilot phase of Golestan cohort study of esophageal cancer. Eur J Clin Nutr 2006;60:971-7.
- 31. Morrow WJ, Ohashi Y, Hall J, Pribnow J, Hirose S, Shirai T, et al. Dietary fat and immune function. I. Antibody responses, lymphocyte and accessory cell function in (NZBxNZW) F1 mice. J Immunol 1985;135:3857-63.
- 32. Lin BF, Huang CH, Chiang BL, Jeng SJ. Dietary fat influences ia antigen expression, cytokines and prostaglandin E2 production of immune cells in autoimmune-prone NZBxNZW F1 mice. Br J Nutr 1996;75:711-22.
- Lin BF, Jeng SJ, Chiang BL, Huang CC. Dietary fat affects lipids and anti-cardiolipin antibody levels in autoimmune-prone NZB/W F1 mice. Br J Nutr 1997;77:657-69.
- Hwang D. Essential fatty acids and immune response. FASEB J 1989;3:2052-61.

Derakhshande-Rishehri, et al.: Fatty acids intake and vitiligo

- 35. Stossel TP, Mason RJ, Smith AL. Lipid peroxidation by human blood phagocytes. J Clin Invest 1974;54:638-45.
- Gerzer R, Brash AR, Hardman JG. Activation of soluble guanylate cyclase by arachidonic acid and 15-lipoxygenase products. Biochim Biophys Acta 1986;886:383-9.
- McPhail LC, Clayton CC, Snyderman R. A potential second messenger role for unsaturated fatty acids: Activation of Ca2+-dependent protein kinase. Science 1984;224:622-5.
- Park S, Lim Y, Shin S, Han SN. Impact of Korean pine nut oil on weight gain and immune responses in high-fat diet-induced obese mice. Nutr Res Pract 2013;7:352-8.
- Jahromi SR, Toghae M, Jahromi MJ, Aloosh M. Dietary pattern and risk of multiple sclerosis. Iran J Neurol 2012;11:47-53.
- Ghadirian P, Jain M, Ducic S, Shatenstein B, Morisset R. Nutritional factors in the aetiology of multiple sclerosis: A case-control study in montreal, Canada. Int J Epidemiol 1998;27:845-52.
- Ghorbanibirgani A, Khalili A, Rokhafrooz D. Comparing *Nigella sativa* oil and fish oil in treatment of vitiligo. Iran Red Crescent Med J 2014;16:e4515.

- Kaimal S, Thappa DM. Diet in dermatology: Revisited. Indian J Dermatol Venereol Leprol 2010;76:103-15.
- Lands WE, Letellier PR, Rome LH, Vanderhock JY. Inhibition of prostaglandin biosynthesis. Adv Biosci 1973;9:15-28.
- Hwang DH, Boudreau M, Chanmugam P. Dietary linolenic acid and longer-chain n-3 fatty acids: Comparison of effects on arachidonic acid metabolism in rats. J Nutr 1988;118:427-37.
- 45. Ochi K, Yoshimoto T, Yamamoto S, Taniguchi K, Miyamoto T. Arachidonate 5-lipoxygenase of guinea pig peritoneal polymorphonuclear leukocytes. Activation by adenosine 5'-triphosphate. J Biol Chem 1983;258:5754-8.
- Millar JH, Zilkha KJ, Langman MJ, Wright HP, Smith AD, Belin J, et al. Double-blind trial of linoleate supplementation of the diet in multiple sclerosis. Br Med J 1973;1:765-8.
- Bates D, Fawcett PR, Shaw DA, Weightman D. Polyunsaturated fatty acids in treatment of acute remitting multiple sclerosis. Br Med J 1978;2:1390-1.
- Paty DW, Cousin HK, Read S, Adlakha K. Linoleic acid in multiple sclerosis: Failure to show any therapeutic benefit. Acta Neurol Scand 1978;58:53-8.

